## A New Validated Stability Indicating RP-HPLC Method for Estimation of Vorasidenib in Bulk and in its Pharmaceutical Dosage Form

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## Abstract

For the estimation of Vorasidenib in bulk and in its pharmaceutical dosage form, a reversed phase high-performance liquid approach chromatographic has been designed and validated in the current work. The separation of Vorasidenib was achieved on Waters Alliance-e2695 by using an X-Bridge Phenyl column (250x4.6mm, 5µ) by eluting with a mobile phase consisting of a mixture of acetonitrile and 0.1% Trifluoroacetic acid in the ratio of 40:60v/vat a flow rate of 1.0 mL/min; detection was carried out by absorption at 234 nm using a photodiode array detector at ambient temperature. The total run time set for the elution of the compound was 5 min. Under the optimised chromatographic conditions, the retention time was obtained at 2.855 min. The current analytical technique validation was conducted in accordance with ICH standards (ICH, Q2R1). The concentration range forVorasidenib in the linearity study was found to be 20-120 µg/mL and the coefficient of variance was found to be 0.9999. The percentage recovery was found to be 99.6-100.3%. LOD and LOQ were found to be 0.48 µg/mL and 1.6 µg/mL respectively. The developed method was also applied to monitor the forced degradation studies on the drug for testing for its ability to resolve the drug from their degradation products. The specificity of the developed method was evaluated by applying acid, base, oxidation, thermal, photolytic and neutral stress conditions to the drug. It was concluded that the estimation of Vorasidenib in bulk and its pharmaceutical dosage form was found to be successfully conducted by using the method.

**Keywords:** RP-HPLC, PDA Detector, Vorasidenib, Method Validation

## Introduction

Vorasidenib (Figure 1) is a targeted cancer drug that treats astrocytoma or oligodendroglioma types of brain tumours in adults and children. The drug inhibits the enzymes of Isocitrate dehydrogenase-1(IDH1) & Isocitrate dehydrogenase-2 (IDH2). It is available in 20 & 40mg of oral tablet formulation. The available brand is Voranigo. Chemically it is 6-(6-Chloro pyridine-2-yl]-N2, N4-bis[(2R)-1,1,1-trifluoro propan-2yl]-1,3,5-triazine-2,4-diamine. It's an empirical formula  $C_{14}H_{13}CIF_6N_{68}$ and molecular weight of 414.74g/mol (1-10).

A literature survey revealed that only one analytical method has been reported for estimating Vorasidenib in bulk and its pharmaceutical dosage form. The reported method is RP-HPLC (11). The present study aimed to develop a simple, sensitive, rapid, and precise RP-HPLC method for estimating Vorasidenib. The analytical method was validated according to ICH validation parameters.

## Materials and Methods

## Chemicals and reagents

A pure sample of Vorasidenib was obtained from Servier India Private Limited, Mumbai. The marketed formulation of

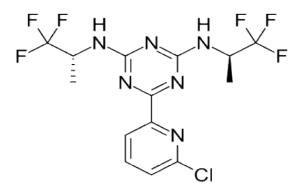


Figure 1: Chemical structure of Vorasidenib

Vorasidenib (Voranigo 40 mg/tablet) was purchased from local pharmacy store. Acetonitrile (HPLC grade) was procured from Rankem®, India, Trifluoroacetic acid (HPLC grade) was procured from Thermo Fisher Scientific India Pvt. Ltd., Mumbai, India, and HPLC graded water and methanol were purchased from Merck Specialties Pvt. Ltd, Mumbai, India. The study made use of water that has been purified using a Milli-Q system.

#### Instrumentation

The analysis was performed by using a chromatographic system, Waters alliance HPLC system, Quaternary gradient pump of Waters Alliance-e2695 series equipped with an auto sampler injector with 10µL is injector loop by using a photo diode array detector and running on E-Z Chrome software with a reverse phase by using Phenyl column 250 x 4.6 mm internal diameter, 5µm particle size. UV-Visible Spectrophotometer (LAB INDIA-3200+), Shimadzu electronic balance (AX-200) was used for weighing purpose. Ultrasonicator (Citizen) and Class "A" volumetric glassware were employed for the study.

#### Chromatographic conditions

Vorasidenib was analyzed with Phenyl column (250x4.6mm, 5µ Particle size) for the chromatographic separation and column was maintained at ambient temperature. The mobile phase was composed of a mixture of Acetonitrile and0.1% Trifluoro acetic acid in the ratio of 40:60% v/vand it was delivered at a flow rate of 1.0 mL/min and detection was monitored at 234 nm with PDA detector. Mobile phase was used as diluent. Injection volume was 10  $\mu$ L. The run time was 5 min. The retention time of Vorasidenib was found to be 2.855 min.

#### Preparation of TFA buffer solution

One millilitre of tri fluoro acetic acid is dissolved in one litre of HPLC graded water and filtered through 0.45µ nylon syringe filter.

### Preparation of mobile phase

Mobile phase was prepared by mixing 0.1% tri fluoro acetic acid and acetonitrile taken in the ratio 60:40. It was filtered through  $0.45\mu$  membrane filter to remove the impurities which may interfere in the final chromatogram.

#### Diluent

Acetonitrile is used as a diluent.

#### Preparation of standard solution

Accurately weighed and transferred 8mg of Vorasidenib working standard into a 10 mL clean and dry volumetric flask and diluent was added and sonicated to dissolve it completely and made volume up to the mark with the same solvent to get Stock solution. Further pipetted 1mL of the above stock solutions into a 10 mL volumetric flask and diluted up to the mark with diluent to get 80ppm of Vorasidenib.

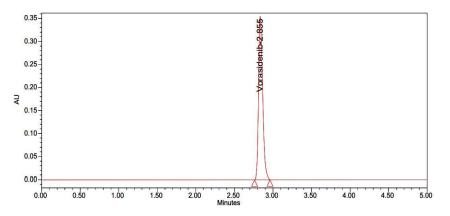


Figure 2: Chromatogram of Vorasidenib standard

## Sample Solution Preparation

Accurately weighed and transferred an equivalent to 18.8mg of Vorasidenib sample into a 10mL clean dry volumetric flask diluent was added and sonicated for 30 min to dissolve it completely and made volume up to the mark with the same solvent. Then it is filtered through 0.45microninjection filter. Further pipetted 1 mL of the above stock solutions into a 10mL volumetric flask and diluted up to the mark with diluent to get 80ppm of Vorasidenib.

## Method Development and Optimization

Several mobile phases of various compositions were examined to provide an optimization of chromatographic conditions for system suitability such as retention time, tailing factor, peak resolution, and theoretical plate count etc. Initially, a mixture of formic acid and acetonitrile in ratios of 20:80, 30:70. and 40:60 was explored. Additionally, mixtures of 0.1% Trifluoroacetic acid (TFA) and Acetonitrile in the ratios of 50:50 and 40:60 were also tested. The optimization process included adjusting the flow rates and mobile phase ratios to achieve the best separation. Ultimately, the mobile phase consisting of 0.1% TFA and acetonitrile (60:40, v/v) at a flow rate of 1.0 mL/min was found to provide the most satisfactory chromatographic performance. This system resulted in well-resolved peaks with good shape and was selected as the optimal

method for the determination of Vorasidenib both in bulk and in its pharmaceutical dosage form.

### Method Validation

The method was validated for Specificity, linearity, accuracy, precision, limit of detection, limit of quantification and robustness by following procedures as per ICH guidelines (12).

#### **Results and Discussion**

The developed method was optimised for the determination of Vorasidenib in bulk and in its pharmaceutical dosage form. Resulted in peak with good shape and well resolved. The results of the optimized HPLC conditions were shown in Figure 2 and Table 1.

The proposed method was validated according to the ICH Q2R1 guidelines which include system suitability, specificity, linearity, accuracy, precision, limit of detection (LOD), limit of quantification (LOQ) and robustness.

## System suitability

In the present study, parameters such as plate count (N), tailing factor (T), resolution (Rs) and reproducibility (%RSD) are determined from replicate injection of standard solution. The acceptable limit of %RSD is less than 2%. Table 2 shows that system suitability results obtained in the present study for Vorasidenib.

	Optimized chromatographic			
conditions				
Parameter	Observation			
Instrument	Waters Alliance e-2695			
used	HPLC			
Column	Waters X-Bridge Phenyl			
	(250x4.6 mm, 5 µm)			
Mobile	Acetonitrile and 0.1% Tri-			
Phase	fluoro acetic acid (40:60)			
Flow Rate	1 mL/min			
Runtime	5 min			
Injection	10 µL			
volume	-			
Detection	234 nm			
Wavelength				
Temperature	Ambient (25°C)			
Mode of	Isocratic mode			
separation				

Table 2: System suitability parameters								
for Vorasidenib								
S.	Parameter	Vorasidenib						
No.								
1	Retention time (min)	2.855						
2	Plate count	11958						
3	Tailing factor	0.90						
4	Resolution							
5	%RSD	0.21						

## Specificity

The specificity of an analytical method is to determine the effect of excipients and other additives that are generally present in the formulation. In this study, the method was evaluated by injecting 10 µL of placebo, standard solution and sample solution into the HPLC system. The test results obtained were contrasted with the results of the standard drug. The chromatograms were checked for the interference peaks shown in Figure 3. Retention time of Vorasidenib wasfound 2.855 min. We did not find any interfering peaks in blank and placebo at retention times of these drug in this method. So, this method was said to be specific.

## Linearity and Range

The linearity of the method was determined at six concentration levels

Table 3: Results of Linearity for Vorasidenib						
	Vorasidenib					
S. No.	Concentration	Peak				
	(µg/mL)	area				
1	20.00	842684				
2	40.00	1569102				
3	60.00	2334947				
4	80.00	3158539				
5	100.00	3934065				
6	120.00	4784103				
Regression	y = 39558.05 x+	4204 14				
equation	y – 39000.00 X+	4294.14				
Slope	39558.05					
Intercept	4294.14					
$R^2$	0.9999					
ronging from	E0 120 ug/ml fo	r the drug				

ranging from 50-120 µg/mL for the drug. Evaluation of the drug was performed with a PDA detector at 234 nm, peak area was recorded for all the peaks. The linearity of the method was evaluated by linear regression analysis. The results shown that an excellent correlation exists between peak area and concentration of drugs within the concentration range indicated. Linear regression data and linearity curves for both analytes were given in Table 3 and Figure 4.

## Accuracy

The accuracy of the method was evaluated by standard addition method. Known amounts of the reference standard solution were added at three different concentration levels to the working standard solution of the drug. The solutions were analysed (n=3) and the mean recovery of the drug at each concentration level was computed and the %RSD was calculated. The studies were performed for Vorasidenib at three different concentration levels (50, 100 and 150%). 10 µL of the samples were injected into the HPLC and the percent recovery and percent RSD were calculated. The corresponding results obtained in the experiments were shown in Table 4.

## Precision

In this study, the precision of the method is determined as Intra-day precision and Inter-day precision. The former is determined by injecting 10  $\mu$ L of the sample for

six times and noting the peak areas of the analytes, taking the average value and finding out the percent RSD. Similarly, the inter-day precision is determined by injecting the sample six times each on two consecutive days, taking the overall average value and calculating the %RSD. The results of the above parameters obtained are summarized in Table 5.

#### Robustness

It is the capacity of a method to remain unaffected by small deliberate variations in method parameter values. The relevant values obtained in the present study by changing the flow rate, mobile phase composition is presented in Figure 5 and Table 6.

# *Limit* of *Detection* and *Limit* of *Quantification:*

The LOD and LOQ values of Vorasidenib from standard deviation of the response and the slope values obtained from their linearity curve, LOD and LOQ for

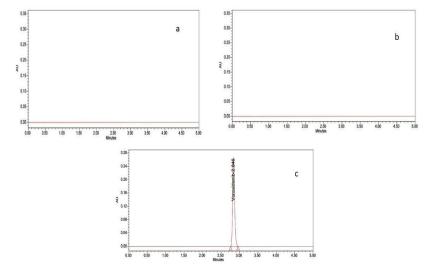


Figure 3: (A) Chromatogram of Blank; (B) Chromatogram of Placebo; (C) Optimized chromatogram

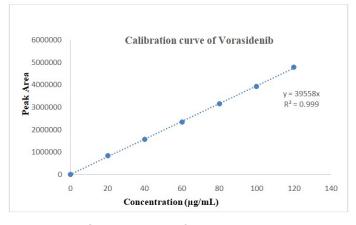


Figure 4: Calibration curve for Vorasidenib at 234 nm Sri et al

Table 4: Accuracy results of Vorasidenib								
Concentration (at specification Level)	Area	Amount Added (mg)	Amount Found (mg)	% Recovery	Mean %Recovery			
	1586487	4.0	4.02	100.5				
50%	1574096	4.0	3.98	99.5	100.3			
	1593432	4.0	4.03	100.8				
	3165234	8.0	8.01	100.1				
100%	3130265	8.0	7.92	99.0	99.6			
	3148796	8.0	7.97	99.6				
	4722130	12.0	11.95	99.6				
150%	4698521	12.0	11.89	99.1	99.7			
	4758103	12.0	12.04	100.3				

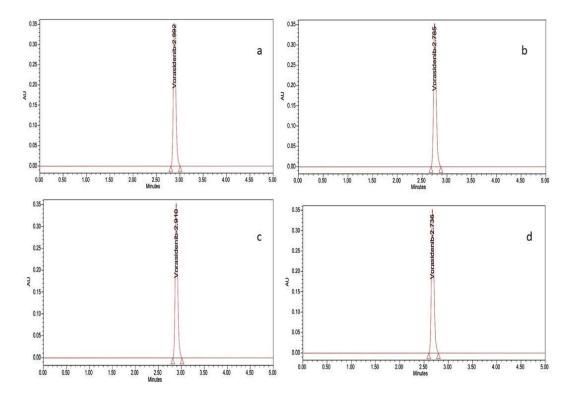
	Table 5: Precision study results of Vorasidenib							
	Concentration	System Precision	Method Precision					
S. No.	Vorasidenib (μg/mL)	Peak area	Peak area					
1	80	3161273	3145206					
2	80	3149174	3178521					
3	80	3165906	3180740					
4	80	3155689	3162963					
5	80	3163622	3140687					
6	80	3166453	3158842					
	Mean	3160353	3161160					
	S. D 6727.58		16535.287					
	%RSD	0.213	0.52					

Vorasidenib was shown in Figure. 6 and Table 7, Which shows the sensitivity of the method.

# Estimation of drug in the tablet dosage form:

The proposed validated method was applied for the determination of Vorasidenib in commercial formulations. The % assay was found to be 99.5 %, Table 8 depicts the assay's results.

*Forced degradation studies on the drugs:* In the present study, forced degradation studies were carried out by subjecting the mixed standard solutions of Vorasidenib to acidic (1N HCl), alkali (1N NaOH), oxidative (3% H<sub>2</sub>O<sub>2</sub>), dryheat (60<sup>o</sup>C/ 6h), photolytic (UV chamber/7 Days) and Neutral degradation (HPLC grade water for 6 h at 60°C) conditions in a laboratory setting to evaluate stability, and the results and data are compared with standard chromatograms. The results obtained from the above experiments indicate that certain amount of degradation of the drug was observed in case of acid and alkaline, peroxide and thermal stress conditions, whereas small extra peaks were found in the relevant chromatograms. It also observed that the purity angle was always less than the purity threshold and it indicates the proposed method was homogenous and can identify the degraded peaks. The chromatograms and drug degradations data obtained in the study were shown in Figure 7 and Table 9.



**Figure 5:** (A) Chromatogram for less flow rate (0.9mL); (B) Chromatogram for more flow (1.1 mL); (C) Chromatogram for less Organic Phase (36:64); (D) Chromatogram for more Organic Phase (44:56)

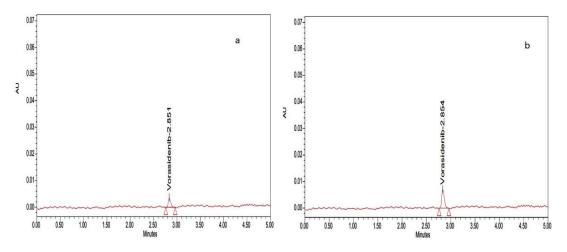
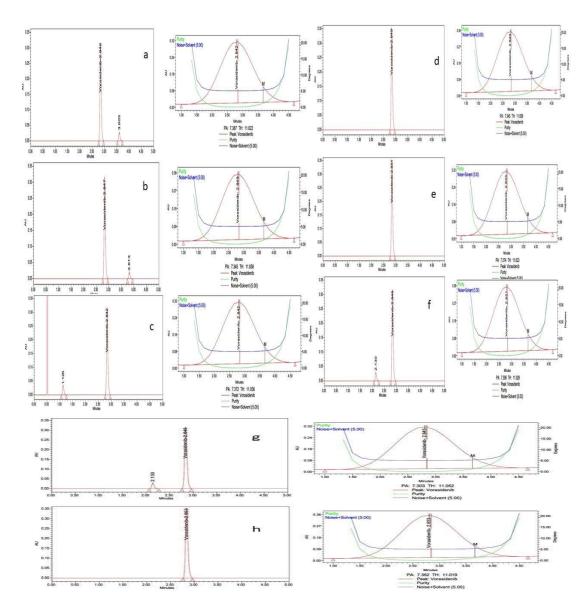


Figure 6: (A) Chromatogram for LOD; (B) chromatogram for LOQ



**Figure 7:** Forced Degradation study chromatograms of Vorasidenib. (A) Chromatogram of acid degradation and Purity Plot of Acid degradation; (B) Chromatogram of alkali degradation and Purity Plot of Alkali degradation; (C) Chromatogram of peroxide degradation and Purity plot of reduction degradation and Purity plot of reduction degradation; (E) Chromatogram of hydrolysis degradation and Purity plot of hydrolysis degradation; (F) Chromatogram of control degradation and Purity plot of control degradation; (G) Chromatogram of thermal degradation and Purity Plot of thermal degradation; (G) Chromatogram of thermal degradation and Purity Plot of thermal degradation; (G) Chromatogram of thermal degradation and Purity Plot of thermal degradation; (H) Chromatogram of photolytic degradation and Purity Plot of photolytic degradation photolytic degradation and Purity Plot of photolytic degradation phot

Table 6: Robustness results of Vorasidenib by HPLC								
Parameter	Condition	Condition Retention time (min) Peak area		Tailing	Plate count			
Elow roto	Less flow (0.9 mL)	2.892	3111575	0.98	12053			
Flow rate Change (mL/min)	Actual flow (1 mL)	2.855	3161273	0.90	11958			
	More flow (1.1 mL)	2.785	3170798	0.86	11871			
Organic	Less Org. (36:64)	2.910	3127258	0.95	12097			
Phase change	Actual (40:60)	2.847	3149174	0.92	11962			
	More Org. (44:56)	2.736	3196341	0.83	11824			

Table 7: Sensitivity parameters (LOD & LOQ) by HPLC							
Name of drug LOD (µg/mL) S/N LOQ (µg/mL) S/N							
Vorasidenib							

	Table 8: Assay results of Vorasidenib								
Brand	Drug	Area	Average	Std.	Sample	Label	Std	Amount	%
			sample	Wt.	wt.	amount	purity	found	assay
			area	(mg)	(mg)	(mg)		(µg/ml)	
Vorani	Vorasi	3139682	3143133	8	18.8	40	99.98	7.96	99.5
go	denib	3146584	5145155	0	10.0	40	99.90	7.90	99.5

	Table 9: Forced Degradation results for Vorasidenib									
Condition W	Sample	Area Counts Injections	Mean Area Count	%	Purity	Purity	%			
	Weight In mg			Label Claim	Angle	Threshold	Degra- dation	Pass/Fail		
Control	18.8	3159154	3159154	100	7.395	11.028	0	Pass		
Acid	18.8	2775423	2775423	87.9	7.387	11.022	12.1	Pass		
Alkali	18.8	2820913	2820913	89.3	7.380	11.035	10.7	Pass		
Peroxide	18.8	2705036	2705036	85.6	7.372	11.036	14.4	Pass		
Reduction	18.8	3096834	3096834	98.0	7.345	11.058	2.0	Pass		
Thermal	18.8	2758611	2758611	87.3	7.303	11.052	12.7	Pass		
Photolytic	18.8	3035306	3035306	96.1	7.362	11.019	3.9	Pass		
Hydrolysis	18.8	2810105	2810105	89.0	7.374	11.023	11.0	Pass		

These outcomes showed that the suggested method is specific and sensitive for quantifying Vorasidenib.

#### Conclusion

The developed method is simple, precise, accurate and reliable for the

estimation of Vorasidenib in Pharmaceutical dosage form. The %RSD of all results is less than 2% that shows high degree. Hence, the proposed method is specific and cost-effective and can be used for routine quantitative analysis of Vorasidenib in its Pharmaceutical dosage form.

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## **Conflict of Interest**

The authors declare that no conflict of interest.

## References

1. https://www.accessdata.fda.gov/scripts /cder/daf/index.cfm?event=BasicSearch.proc ess accessed in on 22<sup>nd</sup> January 2024.

2. Mellinghoff, I.K., Peters, K.B., Cloughesy, T.F., Burris, H.A. III, Maher, E.A., Janku, F., Cote, G.M., de la Fuente, M., Clarke, J.L., Le, K., Guo, R., Yuen, M., Arnofsky, M., Hassan, I., Steelman, L., Pandya, S.S. and Wen, P.Y. (2020). Vorasidenib (VOR; AG-881), an inhibitor of mutant IDH1 and IDH2, in patients with recurrent/progressive glioma: Updated results from the phase I nonenhancing glioma population. *The New England Journal of Medicine*, 29(31), pp.2504-2518.

3. Mellinghoff, I.K., Penas-Prado, M., Peters, K.B., Burris, H.A. III, Maher, E.A., Janku, F., Cote, G.M., de la Fuente, M.I., Clarke, J.L., Ellingson, B.M., Chun, S., Young, R.J., Liu, H., Choe, S., Lu, M., Le, K., Hassan, I., Steelman, L., Pandya, S.S., Cloughesy, T.F. and Wen, P.Y. (2021). Vorasidenib, a dual inhibitor of mutant IDH1/2, in recurrent or progressive glioma: Results of a first-in-human phase I trial. *Clinical Cancer Research*, 27(16), pp.4491-4499.

4. Similar, R., Walsh, G., Matalan, R.J., Guziewicz, N. and Perez-Ramirez, B. (2008). Maximizing data collection and analysis during formulation of biotherapeutic proteins. *Bioprocess International*, 6(10), pp.38-45. 5. Wahid, A., Tariq, A., Ahsan, F. and Asif, F. (2023). Vorasidenib: A promising therapeutic breakthrough for diffuse isocitrate dehydrogenase mutant gliomas. *Rare Tumours*, 15(1-2), pp.513-525.

6. Konteatis, Z., Artin, E., Nicolay, B., Straley, K., Padyana, A.K., Jin, L., Chen, Y., Narayaraswamy, R., Tong, S., Wang, F., Zhou, D., Cui, D., Cai, Z., Luo, Z., Fang, C., Tang, H., Lv, X., Nagaraja, R., Yang, H., Su, S.M., Sui, Z., Dang, L., Yen, K., Popovici-Muller, J., Codega, P., Campos, C., Mellinghoff, I.K. and Biller, S.A. (2020). Vorasidenib (AG-881): A first-in-class, brainpenetrant dual inhibitor of mutant IDH1 and 2 for treatment of glioma. *ACS Medicinal Chemistry Letters*, 11, pp.101-110.

7. IUPAC. (1997). Compendium of Chemical Terminology, 2nd ed. (the Gold Book), updated 26 July 2024.

8. IUPAC. (1997). Compendium of Chemical Terminology, 2nd ed. (the Gold Book), updated 26 July 2024.

9. Thomson Reuters. (2015). Journals Ranked by Impact: Toxicology 2014. *Journal Citation Reports,* Web Sciences (Sciences ed.).

10. Van Telling, C. (2007). Pliny's pharmacopoeia or the Roman treat. *Netherlands Heart Journal*, 15(3), pp.118-120.

11. Zakkula, A., Dittakavi, S., Maniyar, M.M. and Syed, N. (2019). Validated HPLC method for simultaneous quantification of mutant IDH1/2 inhibitors (enasidenib, ivosidenib and vorasidenib) in mice plasma: Application to a pharmacokinetic study. *Biomedical Chromatography*, 33(11), pp.1-8.

12. ICH. (2005). ICH Harmonised Tripartite Guidelines. Validation of analytical procedures: Text and methodology Q2(R1). International Conference on Harmonisation of Technical Requirements for Registration of Pharmaceuticals for Human Use, Geneva, pp.1-13.